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The effect of meter-scale lateral oxygen gradients at the sediment-water interface on selected organic matter based alteration, productivity and temperature proxies

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Abstract

A valid assessment of selective aerobic degradation on organic matter (OM) and its impact on OM-based proxies is vital to produce accurate environmental reconstructions. However, most studies investigating these effects suffer from inherent environmental ⁵ heterogeneities. This includes differences in the initial OM composition, as a result of variable upper water column conditions, or from those induced by selective aerobic degradation. In this study, we used surface samples collected along two meter-scale transects and one longer transect in the northeastern Arabian Sea to constrain initial OM heterogeneity, in order to evaluate selective aerobic degradation on temperature, productivity and alteration indices at the sediment-water interface. All of the alteration indices, the higher plant alkane index, alcohol preservation index, and diol oxidation index, demonstrated that they are sensitive indicators for changes in oxygen content at the sediment-water interface. The export production indices, a cholesterol-based

- stanol/stenol and dinoflagellate lipid- and cyst-based ratios, showed significant (more
 than 20%) change over the lateral oxygen gradients. Therefore, they do not exclusively reflect surface water productivity, but can be altered after deposition with varying oxygen content at the sediment-water interface. Two of the investigated proxies, the glycerol dibiphytanyl glycerol tetraethers (GDGT) based TEX₈₆ sea surface temperature indices and a productivity index based on phytol, phytane and pristane, did not
- show any trends related to oxygen concentration at the sediment-water interface. Nevertheless, unrealistic sea surface temperatures were obtained after application of the TEX₈₆, TEX^L₈₆, and TEX^H₈₆ proxies. The phytol-based ratios were likely modified by the sedimentary production of pristane. Our results demonstrate the rapid and selective impact of aerobic organic matter degradation on the lipid and palynomorph composition
- ²⁵ of surface sediments on a small spatial scale and suggests useful tracers of changing redox conditions along the sediment-water interface.



1 Introduction

Organic matter (OM)-based proxies are very useful tools in paleoclimatology and environmental studies of the marine realm. Unfortunately, OM-based proxies can be diagenetically modified, which can result in overprinting of the initial signals and lead to

misleading reconstructions (for a review see Zonneveld et al., 2010). For an adequate interpretation of proxy results, it is therefore essential to have detailed information about the processes responsible for overprinting and the rate at which diagenesis might alter the original signal.

Freshly deposited OM is quickly degraded at the sediment-water interface (SWI) during early diagenesis (Henrichs, 1992; Canuel and Martens, 1996; Prahl et al., 2000; Wakeham et al., 2002). One of the most significant variables influencing OM degradation is the oxygen content in bottom waters and the O₂ exposure time (e.g. Cowie et al., 1995; Hedges and Keil, 1995; Hartnett et al., 1998; Hulthe et al., 1998; Hedges et al., 1999). Furthermore, OM components such as lipid biomarkers (e.g. Hoefs et al., 2002; Sinninghe Damsté et al., 2002; Versteegh et al., 2010) and palynomorphs (e.g. Zonneveld et al., 1997, 2001; Bockelmann et al., 2007; Versteegh et al., 2010), display varying rates of degradation, as some are intrinsically more labile (e.g. Sun and Wakeham, 1994; Wakeham et al., 2002; Zonneveld et al., 2008). This is important

- as biomarkers and palynomorphs are widely used to reconstruct oceanographic conditions, such as productivity (e.g. Schubert et al., 1998; Schulte et al., 1999; Reichart and Brinkhuis, 2003). However, in studies of marine sediments, it is difficult to separate the effects of selective aerobic degradation from other factors, such as differences in upper water column conditions (van der Weijden et al., 1999), wind transported material (Witte and Pfannkuche, 2000), sediment accumulation rates (Hedges and Keil,
- 1995), as well as winnowing (Pedersen et al., 1992), lateral transport (Calvert et al., 1995; Mollenhauer et al., 2007, 2008) and advection, and water depth (Wakeham et al., 2002; Wuchter et al., 2006). These can influence the composition of OM settling on the sea floor and complicate interpretations regarding oxygen effects (Hedges and Keil, 1995).



In this study, we investigate rapid changes in biomarker- and palynomorph-based proxy ratios as a result of laterally increasing oxygen concentrations at the SWI on the Pakistan continental margin (Northeastern Arabian Sea), where the primary control on OM preservation is the bottom water oxygen concentration (e.g. Paropkari et al., 1992; 1993; Cowie et al., 1999; Keil and Cowie, 1999; van der Weijden et al., 1999; Schulte 5 et al., 2000). The higher plant alkane index (HPA; Westerhausen et al., 1993), alcohol preservation index (API; Cacho et al., 2000), and diol oxidation index (DOXI; Ferreira et al., 2001) have previously been used to indicate sediment alteration and oxygenation changes in the sedimentary record. We evaluate whether they are also sensitive indicators of redox changes at the SWI. Additionally, some molecular biomarkers commonly 10 used to indicate changes in export production like phytol, cholesterol, and dinosterol (Volkman et al., 1998) were shown to be more rapidly degraded in an oxygenated environment (e.g. Schulte et al., 2000). In order to determine whether this degradation is selective, we incorporate them into indices along with related refractory compounds,

- ¹⁵ such as pristane and phytane, cholestanol, and dinostanone and dinosterone, respectively (after Nishimura and Koyama, 1977; McCaffrey et al., 1991; Mouradian et al., 2007). Another proxy that is used to show changes in export production is the ratio between autotrophic (gonyaulacoid) and heterotrophic (peridinioid) dinoflagellate cysts (e.g. Harland, 1973; McCarthy et al., 2000; Reichart and Brinkhuis, 2003 and references therein). However, the gonyaulacoid species have been shown to be resistant
- to aerobic degradation while the peridinioid species are sensitive to oxygen exposure (e.g. Zonneveld et al., 1997, 2001; Combourieu-Nebout et al., 1998; Versteegh and Zonneveld, 2002). Here, we compare gonyaulacoid and peridinioid (G/P) dinoflagellate cysts to determine to what extent this ratio may be affected. Finally, we investigate
- ²⁵ marine isoprenoidal archaeal glycerol dibiphytanyl glycerol tetraether (GDGT) distributions (Karner et al., 2001; Wuchter et al., 2005; Menzel et al., 2006), which are the basis for sea surface temperature (SST) proxies based on the number of GDGT cyclopentane moieties (e.g. Schouten et al., 2002; Kim et al., 2008, 2010). There is conflicting evidence on the influence of selective aerobic degradation on GDGTs with



some studies suggesting no selective degradation (e.g. Sinninghe Damsté et al., 2002; Schouten et al., 2004; Kim et al., 2009) while others indicate that differences in lability between the moieties are possible (e.g. Shah et al., 2008; Huguet et al., 2009). In order to provide more information, we investigate the possible effect of a lateral oxygen gradient in surface sediments on GDGT-based indices.

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As the effects of selective aerobic degradation are best studied on samples with similar initial OM compositions, we utilize surface sediment samples collected along three transects that, when used in conjunction, allow us to constrain the uncertainty associated with OM heterogeneity. The first transect encompasses the large, stable, intermediate depth (150–1300 m; e.g. Breuer et al., 2009) oxygen minimum zone (OMZ) and

- ¹⁰ diate depth (150–1300 m; e.g. Breuer et al., 2009) oxygen minimum zone (OMZ) and oxygenated deeper water. Two additional transects include methane seep sites, which are found in this region (e.g. von Rad et al., 1996; Fischer et al., 2011). One seep transect is located at the lower transition of the OMZ and thus basically records the impact of the seep environment on the OM composition. The other seep transect is lo-
- ¹⁵ cated below the OMZ in well-oxygenated bottom waters with oxygen levels increasing with distance from the gas outflow. The active advection of vent fluids at a seep site reduces the oxygen penetration depth and the O₂ concentration at the SWI (Aharon and Fu, 2003). This results in a short scale lateral oxygen gradient extending from just adjacent to the active venting towards the periphery of the seep site. Essentially,
- the meter-scale seep transects serve to constrain environmental variability that may affect OM composition along the longer OMZ transect. Influences on OM contributed by the seep biomass and possible complications from anaerobic degradation (e.g. Canfield et al., 1993; Grossi et al., 2001) via bacterial sulfate reduction (e.g. Jørgensen, 1982; Lückge et al., 1999; Jørgensen and Kasten, 2006) are addressed through the comparison of the seep transects to each other and from the seep transects to the OMZ transect. This study thus provides more detail as to the rapidity of overprinting by changing oxygen content at the SWI on marine surface samples (i.e. assumed short oxygen exposure time) along lateral oxygen gradients.



2 Regional setting

In the study area, two monsoon seasons, the boreal winter northeast monsoon (NEM) and the stronger southwest monsoon (SWM) in the summer, result from land-sea pressure gradients due to differential summer heating of the Asian continent. This produces seasonally reversing wind conditions, which induce high annual primary productivity (Wyrtki, 1973; Qasim, 1982) along the coast and in the open ocean region through upwelling (during SWM) and convective mixing (during NEM) (Bauer et al., 1991; Madhupratap et al., 1996; Prassana Kumar et al., 2001). Abundant OM in the upper water column due to the high primary production leads to a high rate of oxygen consumption as a result of OM degradation. Together with the inflow of O₂-poor intermediate

- waters, these cause a stable, intermediate depth OMZ that has existed for the past 7000 yr (Shapiro and Meschanov, 1991; Olson et al., 1993; von Rad et al., 1999). Current estimates place the OMZ between about 150–1000 m water depth, though there is some variability as to the depths of the upper and lower boundaries (Brand and Grif-
- fiths, 2009; Breuer et al., 2009). The OMZ was defined in this study as having oxygen concentrations below 0.5 ml l⁻¹ (Fig. 1). Average OM export production for the Pakistan continental margin is approximately 70 g C m⁻² yr⁻¹ with a sedimentation rate on the slope of up to 1 mm yr⁻¹ (von Rad et al., 1995, 1999; van der Weijden et al., 1999). Sedimentary OM primarily reflects marine input (e.g. Paropkari et al., 1992; Cowie et al., 1999; Lückge et al., 1999; Schulte et al., 2000), although, there is seasonal ter-
- restrial influx of organic rich mud from the Shadi and Hingol rivers (Bohrmann et al., 2008).

The Makran accretionary prism contains numerous cold seeps, which are areas of fluid and gas emissions from the sea floor (von Rad et al., 1995, 1996). *R/V Meteor* ²⁵ cruise M74/2 and 3 was an interdisciplinary cruise specifically scheduled to explore the Makran continental margin for further fluid escape structures (Bohrmann et al., 2008) driven by tectonically induced overpressure (Ding et al., 2010). Coupled to the upward gas migration at seep sites are chemosynthetic communities that rely on the supply of reduced compounds such as hydrogen sulfide and methane (Suess et al.,



1985). Microbial mats composed of large sulfide-oxidizing bacteria (*Beggiatoa* and *Marithioploca* spp.) and clam fields (*Calyptogena* spp.) associated with such seep sites have been found specifically here (e.g. von Rad et al., 1995, 1996; Bohrmann et al., 2008; Fisher et al., 2011). The biogeography of the seep sites is controlled to some degree by the active fluid and gas expulsion into the water column as well as oxygen availability (Roberts and Carney, 1997; Fischer et al., 2011). Thus, these seep sites can encompass localized areas of anoxia at the SWI in otherwise oxygenated bottom waters (Aharon and Fu, 2000), such as below the OMZ (von Rad et al., 1996). The two seep fields sampled for this study are shown in Fig. 2.

10 3 Material and Methods

3.1 Material

Surface sediments were retrieved along three transects during *R/V Meteor* cruise M74/3 during the 2007 fall intermonsoon (Bohrmann et al., 2008). In the study area, CTD casts identified the lowest oxygen concentrations between 350–750 m water depth with 900–1200 m containing slightly higher O₂ content and well-ventilated waters found below 1600 m (Fig. 1; Table 1). Only the uppermost unconsolidated sediment layer (<1 cm) was used.

3.1.1 OMZ transect

Three multi-cores were retrieved along a kilometer-scale transect starting within the OMZ (GeoB 12312; 0.02 mll⁻¹ O₂; 655 m b.s.l.) into the transition zone (GeoB 12321; 0.55 mll⁻¹ O₂; 1425 m b.s.l.) and extending into the oxic zone below (GeoB 12331; 2.55 mll⁻¹ O₂; 2830 m b.s.l.). This series of samples is hereafter referred to as the OMZ transect (Fig. 1; Table 1), and the samples are tentatively characterized as OMZ-anoxic (GeoB 12312), OMZ-suboxic (GeoB 12321) and OMZ-oxic (GeoB 12331). All sediment samples are composed of clay to silty clay material. They were not retrieved



near areas of active seeping and thus represent background sediments of detrital OMbased ecosystems. OMZ-anoxic contained finely laminated layers and no indications of bioturbation, whereas OMZ-suboxic and OMZ-oxic appeared more homogenized, most likely due to bioturbation (Bohrmann et al., 2008). Subsamples for palynomorph anal-⁵ ysis were sealed air tight and stored at 4 °C until analysis. Subsamples for biomarker analysis were stored under argon and at -20 °C.

3.1.2 Seep transects

Two seep fields (GeoB 12328 and GeoB 12326; Fig. 1; Table 1) were sampled using the remotely operated vehicle (ROV) QUEST 4000 (Bohrmann et al., 2008). This technique allowed us to know the exact push core location as well as view the surrounding environment (Fig. 2). The CTD bottom water oxygen measurements from the two push core stations demonstrated that site GeoB 12328 was located in the transitional OMZ zone (0.07 ml I⁻¹ O₂; 1025 m b.s.l.) and site GeoB 12326 was located in well-oxygenated bottom waters (2.55 ml I⁻¹ O₂; 2875 m b.s.l.) (Fig. 1; Table 1). Subsequently, the two seep fields are referred to as OMZ-seep (GeoB 12328) and below OMZ-seep (GeoB 12326). Three successive push core samples comprise the OMZ-seep transect: GeoB 12328-6 (OMZ-seep 1) was retrieved at the edge of the central bacterial mat, GeoB 12328-2 (OMZ-seep 2) where bubbles of free gas escape at the sea floor at 15 cm distance away and GeoB 12328-4 (OMZ-seep 3) at about 75 cm from

- the active seeping and within the clam field (Fig. 2a; Table 1). As well, three successive push core samples comprise the below OMZ-seep: GeoB 12326-13 (below OMZ-seep 1) was just adjacent to active seeping, GeoB 12326-9 (below OMZ-seep 2) was 50 cm away from the central gas orifice and within the area of small clams, and GeoB 12326-7 (below OMZ seep 3) was 1 m further away at the edge of the small clams and ambient
- sea floor (Fig. 2b; Table 1). When only the seep transect is stated (i.e. OMZ-seep or below OMZ-seep), this refers to all of the push cores retrieved at that respective site. All subsamples for biomarker and palynomorph analysis were stored under argon and at -20 °C.



3.2 Methods

3.2.1 Geochemical parameters

Ex situ pore water oxygen was measured on replicate whole cores of OMZ-anoxic, OMZ-suboxic, and OMZ-oxic with a FIBOX3 oxygen sensor coupled to a micromanipulator. The cores were allowed to acclimate to ambient temperature and then the O_2 5 probe was lowered quickly to the measurement depth and left until the O₂ reading became stable. Measurements were taken at 2.5 mm intervals and in triplicate. This method was primarily used to specify the oxygen penetration depth. The O_2 penetration depth was compared to pore water iron (Fe²⁺) concentrations, as a way to confirm the redox boundary. For this purpose, pore water was extracted on board via rhizons 10 (pore size 0.1 µM) according to the method of Seeberg-Elverfeldt et al. (2005). Subsamples (1 ml) were complexed with 50 µl of "Ferrospectral" and measured photometrically. At the below OMZ-seep, manganese (Mn²⁺) pore water concentrations were measured on two push cores replicate to below OMZ-seep 1 (central habitat, active gas ebullition) and below OMZ-seep 2 (within the clam field). A pore water subsample 15 for cation analysis was diluted 1:10 with 1M HNO₃ and stored at 4 °C. Samples were analyzed for Mn²⁺ concentrations with an ICP-OES (IRIS Intrepid, Thermo Electron). Standards were prepared from single element stock solution in 1M HNO₃⁻ to avoid matrix effects. Replicate measurements gave an error of ≤ 3 %.

20 3.2.2 Lipid biomarkers

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Freeze dried sediment samples (1-10 g) were ground with an agate mortar and pestle, and the total lipid extract (TLE) was extracted using an Dionex 200 ASE (Accelerated Solvent Extractor) with methanol (MeOH) and dicholoromethane (DCM) (9:1 v:v; 3 cycles of 5 min duration each) at 100 °C and 7.6 × 10⁶ Pa. A small aliquot (100 µl) of the TLE was evaporated under N₂ to dryness and then redissolved in 100 µl *n*hexane and propanol (99:1 v:v) for GDGT analysis following the procedure of Liu



et al. (2011), modified after Hopmans et al. (2000). Briefly, a ThermoFinnigan Surveyor high performance liquid chromatography (HPLC) system was used at 30 °C, a flow rate of 1 ml min⁻¹, and separation was achieved with an Econosphere NH₂ column (250 × 4.6 min; Alltech, Germany). The gradient used was the same as Liu et al. (2011). The HPLC was coupled to a ThermoFinnigan LCQ Deca XP Plus ion trap mass spectrometer via an atmospheric pressure chemical ionization (APCI) interface. These settings were also the same as described in Liu et al. (2011). Relative GDGT concentrations were acquired by integrating the appropriate peak areas of their (M + H)⁺ and (M + H)⁺ + 1 mass chromatograms.

- ¹⁰ Another aliquot (500 µl) of the TLE was partitioned into a *n*-hexane soluble fraction and a DCM soluble fraction. The hexane soluble fraction was then derivatized with 50 µl Bis(trimethylsilyl) trifluoroacetamide (BSTFA) and pyridine (60 °C, 1 h) to produce trimethylsilylated alcohols. An internal standard mix (5 µg) containing C_{36} *n*-alkane and 1-nonadecanol was added before derivatization. Samples were then analyzed using an
- Agilent Technologies 7890A Gas Chromatograph (GC) equipped with a splitless injector and a HP-5MS column (i.d. 0.25 mm; film thickness 0.25 μm) operated at a flow rate of 1 ml min⁻¹. Helium was used as the carrier gas. The temperature program was as follows: 70 °C (1 min), then 60–310 °C at 4 °C min⁻¹ and 320 °C (21 min). The GC was coupled to a quadripole mass spectrometer (MS) (597C VL MSD Triple Axis Detec-
- tor) operated at 70 eV and a scan range of 50–800 AMU. Compounds were identified based on relative retention times and indices, and mass spectra. In general, they were quantified by comparing their peak areas in the total ion current with internal standards. The relative response factors were assumed to be 1; hence our data are only semi-quantitative. Since we were interested primarily in the relative changes of proxy ratios
- between samples, this analysis was sufficient. For those compounds where co-elution occurred, quantification was performed on compound specific ions after Versteegh et al. (2010).



3.2.3 Dinoflagellate cysts

Sample material (~ 0.3 g) was oven dried (60 °C; 24 h), weighed and treated with cold 10% HCI (24h) to remove carbonates and 38% HF (48h after 90 min agitation) to remove silicates, then neutralized with 10% KOH. Care was taken to ensure that the solutions never became alkaline. The samples were briefly put in an ultrasonic bath 5 (<1 min) and then sieved through a 20 µm precision sieve (Stork Veco; mesh 317). Samples were centrifuged (3200 rpm; 6 min) and concentrated to 1.0 ml, after which $50\,\mu$ (100 μ for push core samples) were mounted in glycerin jelly and sealed on a glass slide with paraffin wax. Entire slides were counted blind for dinocysts with a Zeiss Axioskop light microscope. At least 200 whole specimens were counted. If nec-10 essary, additional slides were counted entirely. Taxonomy was based on Fensome and Williams (2004) and Matsuoka et al. (2009). Dinoflagellate cyst taxa were divided into two groups, gonyaulacoid (G-cysts) and peridinioid and cysts of Polykrikos (Pcvsts) (Table 2), which generally represent autotrophic and heterotrophic taxa, respectively (e.g. Mudie and Rochon, 2001). Quantification was performed via the volumetric 15 method (e.g. Dale, 1976; Holzwarth et al., 2007).

3.2.4 Proxy ratio calculations

A proxy trend in this study is defined as a consistent increase or decrease along successive samples within a transect. Specific proxy definitions and formulae are de-²⁰ picted in Table 3. The alteration proxy HPA utilizes ΣC_{24-28} even *n*-alcohols divided by the ΣC_{27-31} odd *n*-alkanes (Westerhausen et al., 1993), while the API uses only *n*hexacosanol and *n*-nonacosane (Cacho et al., 2000). The HPA and API ratios exhibit decreasing values if their respective *n*-alcohols are selectively degraded relative to the *n*-alkanes. The final alteration proxy, DOXI, involves the concentration of 1,15-long chain alkyl diols divided by the keto-ols (Ferreira et al., 2001). This proxy assumes that the keto-ols are mainly oxidation products of the diols; thus, higher ratio values indicate higher degradation of the diols relative to the keto-ols. We initially calculated the



DOXI with the dominant C_{30} isomers; the predominance of this isomer was reported previously in Arabian Sea sediments (e.g. Smallwood and Wolff, 2000; Wakeham et al., 2002). Where possible, the ratio was also calculated using the C_{32} isomers.

- For the three phytol-based indices, significant preferential degradation of phytol would result in overall lower ratio values as pristane and phytane are generally considered compounds produced via degradation of phytol (e.g. Didyk et al., 1978). The cholesterol-based stanol/stenol index would show an increase with more efficient cholesterol oxidation. The dinosterol-based index compares dinosterol with other dinoflagellate-derived degradation products, dinosterone, dinostanone, and dinoster-
- ane (Σ-dinoflagellate lipids). Thus, decreases in the proxy would indicate faster removal of dinosterol. Although the original definition of the G/P (gonyaulacoid/peridinioid) ratio is based on the number of species of each (Harland, 1973), in this study we used the quantity of G- and P-cysts in each sample because the small spatial scale of our transects made differences in species numbers highly unlikely.
- ¹⁵ The temperature indices are based on the isoprenoid GDGT distribution. The abundance of different GDGT moieties can be correlated to temperature using the GDGTs with one to three cyclopentane moieties and the crenarchaeol regioisomer, which contains four cyclopentane moieties and one cyclohexane moiety (Wuchter et al., 2004; Schouten et al., 2007). Three indices are calculated based on Kim et al. (2010): TEX₈₆,
- TEX^L₈₆, TEX^H₈₆ (Table 3). From each of these indices, sea surface temperatures (SST) were calculated after Kim et al. (2010), using their linear calibration Eqs (5), (8), and (10) for TEX₈₆, TEX^L₈₆, and TEX^H₈₆, respectively. Changes in the distribution of the GDGTs are thus reflected in the index values and calculated SSTs. All raw data used for the calculation of these indices (lipid biomarker concentrations, GDGT peak areas, and dinoflagellate cyst counts) can be found in a Supplement.



4 Results

4.1 Oxygen content at the SWI

The O₂ penetration depth in the sediments increases along the OMZ transect from undetectable (OMZ-anoxic) to a maximum of about 25 mm (OMZ-oxic), and is consistent with the Fe²⁺ profiles (Fig. 3). At the below OMZ-seep, Mn²⁺ was present at the SWI (~3 µmol I⁻¹) closest to the central habitat of active gas seepage (below OMZ-seep 1), but concentrations were lower (~0.4 µmol I⁻¹) 50 cm away at the SWI in the clam field (below OMZ-seep 2) (Fig. 4).

4.2 Proxy ratios

10 4.2.1 Alteration indices (HPA, API, DOXI)

Along the OMZ transect, HPA (Table 3) values decreased from 0.58 (OMZ-anoxic) to 0.42 (OMZ-oxic). Along the below OMZ-seep, the index decreased from 0.67 (below OMZ-seep 1) to 0.55 (below OMZ-seep 3). At the OMZ-seep, the index values ranged from 0.6–0.73 with no apparent trend.

¹⁵ Along the OMZ transect, the API (Table 3) values decreased from 0.47 (OMZ-anoxic) to 0.18 (OMZ-oxic). Additionally, at the below OMZ-seep, the ratio values decreased from 0.56 to 0.37 with increasing distance from the active venting. Values at the OMZ-seep were higher in general (0.61–0.72), but showed no specific trend (Table 4).

The DOXI (Table 3) increased along the OMZ transect from 0.12 (OMZ-anoxic) to 0.53 (OMZ-oxic). Along the below OMZ-seep, DOXI also increased from 0.26 (below OMZ-seep 1) to 0.41 (below OMZ-seep 3). At the seep station within the OMZ, DOXI varied between 0.25–0.29. When it was possible to also calculate the ratio using the C_{32} 1, 15-diol and keto-ol, values were similar to the C_{30} ratios (Table 4).



4.2.2 Export production proxies (phytol, cholesterol, dinosterol indices, and dinoflagellate cysts)

Along the OMZ transect, phytol indices (Table 3) containing pristane, i.e. index¹ and index³, remained around 0.25 and 0.15, respectively (Table 4). Phytane-containing index² values were between 0.08–0.16 and showed no apparent trend. At both the OMZ-seep and below OMZ-seep, ratio values were generally higher overall, especially in the samples closest to active venting (Table 4). Below the OMZ-seep, all index values showed a decrease. Index¹ decreased from 0.51 (below OMZ-seep 1) to 0.25 (below OMZ-seep 3), index² from 0.2 to 0.1, and index³ values decreased from 0.32

to 0.15. Values of index¹ at the OMZ-seep began extremely high (1.26; OMZ-seep 1) and decreased to 0.47 (OMZ-seep 3) while index³ decreased from 0.99 to 0.32. Index² showed less of a decrease from 0.28 to 0.15 (Table 4).

The cholesterol-based stanol/stenol ratio (Table 3) showed an increase along the OMZ transect from 0.23 (OMZ-anoxic) to 0.82 (OMZ-oxic). An increasing trend (0.37

to 0.75) was also apparent along the seep transect below the OMZ. Along the OMZseep, values ranged between 0.4–0.61 (Table 4).

The dinosterol-based index values increased along the OMZ transect (0.02 to 0.24) and the below OMZ-seep (0.03 to 0.26), and decreased along OMZ-seep (0.18 to 0.06) (Table 4).

²⁰ The G/P ratio derived from dinoflagellate cysts (Table 3) increased from 0.13 (OMZanoxic) to 0.52 (OMZ-oxic) along the OMZ transect and from 0.15 to 0.44 along the below OMZ-seep. The ratio was consistent (0.17–0.18) at the OMZ-seep.

4.2.3 Temperature proxy

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The TEX₈₆ index decreased slightly from 0.75 (OMZ-anoxic, OMZ-suboxic) to 0.72 (OMZ-oxic) along the OMZ transect (Table 4). The corresponding SSTs showed a decrease from 34.6 to 31.7° C. TEX₈₆ also decreased along the below OMZ-seep from 0.76 (below OMZ-seep 1) to 0.69 (below OMZ-seep 3). The calculated SSTs



decreased from 35.6 to 29.2 °C. Along the OMZ-seep, TEX₈₆ ranged from 0.71–0.75, with resulting SSTs between 31.0–34.9 °C. TEX₈₆ showed no clear trends along any transect and the index values ranged from -0.33 (below OMZ-seep 2 and 3) to -0.28 (OMZ-seep 3). Subsequent SSTs varied from 24.3 °C (below OMZ seep-2) to 28.3 °C

⁵ (OMZ-seep 3). Along the OMZ transect and below OMZ-seep, TEX^H₈₆ values slightly decreased from -0.12 to -0.15 and -0.12 to -0.16, respectively. This resulted in SSTs for the two transects as 30.1–28.6 °C, and 30.6–27.4 °C. The OMZ-seep showed a slight increase (-0.15 to -0.12) and subsequent increase (28.7 to 30.3 °C) in TEX^H₈₆ and SST, respectively.

10 **5 Discussion**

5.1 Lateral oxygen gradients

Two of the investigated transects are characterized by differences in the oxygen content at the SWI and thus have the potential to exert a strong diagenetic overprint on our proxy ratios. The oxygen gradient through the OMZ is clear in our data (Fig. 3)
and well known from other studies (e.g. Breuer et al., 2009). This confirms our initial classification (Table 1) based on CTD profiles of the study area (Fig. 1b) and sediment descriptions of the OMZ transect multicores (Bohrmann et al., 2008). Although there was a very small amount of oxygen measured in the bottom waters at OMZ-anoxic, we classified it as such in order to distinguish it from OMZ-suboxic, for which the bottom
water oxygen concentration is one order of magnitude higher. This same logic applies to the classification of the OMZ-seep, located at the lower transition of the OMZ. The bottom water O₂ measurement of the station (0.07 mII⁻¹; Fig. 1b) is very close to that of OMZ-anoxic, so all samples were classified as anoxic (Table 1). However, we should note that while the OMZ-seep central habitat contained a microbial mat, indicat-

ing anoxia, the presence of small vesicomyid clams further away from the gas orifice does suggest that O₂ concentrations were high enough to support them (e.g. Fischer



et al., 2011). Therefore, it is likely that a lateral oxygen gradient was present at the OMZ-seep; however, the magnitude of this gradient must be smaller than along the OMZ transect or the below OMZ-seep.

- The presence of a distinct spatially short-scale oxygen gradient at the below OMZseep was inferred from the differences in fluid/gas flow and colonization by chemosynthetic communities of the seep habitats (Bohrmann et al., 2008; Fischer et al., 2011). The pore water manganese (Mn²⁺) profiles (Fig. 4) corroborate this, indicating oxygen was either absent or present in very low concentrations at below OMZ-seep 1 and higher at the SWI of below OMZ-seep 2. Mn-oxides are stable under oxic conditions and begin to dissolve as soon as the environment becomes reducing (Saager et al., 1989; Schenau et al., 2002; van der Weijden et al., 2006). So, the presence or absence of Mn²⁺ in the uppermost sediments can be used as an oxygen indicator. Subsequently, the below OMZ-seep 2 as suboxic, and below OMZ-seep 3 as oxic (Table 1).
- ¹⁵ Below OMZ-seep 3 was classified as oxic since it is furthest from the central gas orifice and more likely influenced by the bottom water oxygen content of the surrounding area.

5.2 Alteration proxies (HPA, API, DOXI)

5.2.1 HPA

In the transects incorporating lateral oxygen gradients, the HPA ratios decrease by
 almost 30% (OMZ transect) and 20% (below OMZ-seep) with increasing oxygen concentrations (Fig. 5a). In comparison, there was no change observed along the OMZ-seep (Fig. 5a). These trends are the result of preferential degradation of the *n*-alcohols, which are more labile than the *n*-alkanes (e.g. Yamamoto et al., 2008). Since these trends are similar along the long OMZ transect and the short below OMZ-seep,
 the ratio values cannot only be a result of degradation en route to the marine realm or during settling through the water column, which confirms the conclusions of West-

or during settling through the water column, which confirms the conclusions of Westerhausen et al. (1993). The similar HPA values along both lateral oxygen gradients



indicate that the major factor affecting this ratio is the oxygen content, which indicates that this index may be a useful and sensitive proxy to determine redox changes at the SWI.

5.2.2 API

⁵ The API provides very clear evidence for selective aerobic degradation at the SWI (Fig. 5a). The index decreased by more than 60 % along the OMZ transect and more than 30 % along the below-OMZ seep, whereas no trend was visible along OMZ-seep. In the studies of Cacho et al. (2000) and Martrat et al. (2007) where this ratio was used to reflect bottom water oxygenation changes in sediment cores, environmental heterogeneity, such as fluctuations in terrestrial input over time, could not be discounted as a variable possibly affecting the proxy. The short lateral oxygen transect of below OMZ-seep effectively eliminates that uncertainty. In our data, this ratio is effective as a bottom water oxygen indicator and is sensitive to changes in oxygen content at the SWI. Thus, we suggest that it is also a possible proxy for oxygenation changes in surface samples.

5.2.3 DOXI

This index reflects significant and rapid selective aerobic degradation, as a five-fold increase is present in the 1,15-C₃₀ index along the OMZ transect and a doubling along the below OMZ-seep (Fig. 5a). Selective degradation of the diols relative to the keto-

- ols has previously been used to distinguish between the oxidized and unoxidized portions of the Mediterranean S1 sapropel (Ferreira et al., 2001; Versteegh et al., 2010). However, this is the first indication that this proxy is also able to reflect redox changes at the SWI in freshly deposited material. Ferreira et al. (2001) noted that the absolute concentrations of keto-ols increased as the diol concentrations decreased. They
- ²⁵ proposed that the keto-ols could be an oxidative product of the diols. In our samples, the relative concentration of the keto-ol increased as well (see Supplement), which is



further evidence that keto-ols are oxidation products of the diols. C_{28} – C_{32} keto-ols were identified in a cultured marine eustimatophyte, *Nannochloropsis gaditana* (Méjanelle et al., 2003) and a fern species, *Osmunda regalis* (Jetter and Riederer, 1999). However, the keto-ol concentrations reported in *N. gaditana* were less than what is generally measured in marine sediment (Méjanelle et al., 2003). Likewise, the terrestrial input to our samples is much less than the marine component so influence from vegetation should not be a major factor. Thus, even if there is a contributing factor of biosynthesized keto-ols, it is most likely a small amount and still would not be sufficient to explain the trend in the ratios along the seep transect below the OMZ.

10 5.3 Export production proxies (phytol, cholesterol, dinosterol indices and dinoflagellate cysts)

5.3.1 Phytol-based indices

The lack of any significant trends attributable to changing oxygen content from the three phytol-based indices indicates they do not reflect selective aerobic degradation. Instead, a secondary source appears to be overprinting two phytol-based indices. Index¹ 15 and index³ (Table 4; Fig. 5b) values in below OMZ-seep 1 and OMZ-seep 1 are up to an order of magnitude higher in comparison to the non-seep influenced anoxic sample, OMZ-anoxic. In contrast, index² values only vary between 0.1–0.28 in all samples (Fig. 5b). This may suggest a sedimentary contribution of pristane. While pristane and phytane are generally considered compounds produced via degradation of phytol 20 (e.g. Didyk et al., 1978), this assumption for pristane has been questioned in anoxic sediments (e.g. Grossi et al., 1998; Rontani and Volkman, 2003; Rontani et al., 2010). The higher index¹ and index³ values in below OMZ-seep 1 and OMZ-seep 1 could indicate faster conversion of reactive phytol into a more refractory form (i.e. bound), which dominates in anoxic settings (e.g. Sun et al., 1998). If this were true, it should also be 25 reflected in index², where in fact the values are stable. Thus, it is likely that there is a secondary source of pristane that does not involve the degradation of chlorophyll-a (for



a review see Rontani and Bonin, 2011). Sources could include tocopherol degradation (Rontani et al., 2010) or input from archaeal lipids (Rowland, 1990). Even without the sedimentary input of pristane, the phytane-containing index² does not show preferential degradation of phytol with increasing SWI oxygen concentrations. Our results agree with previous studies cautioning against the use of a proxy for redox changes based on these assumed phytol degradation products (Rontani and Volkman, 2003).

5.3.2 Cholesterol based stanol/stenol index

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Increasing cholestanol/cholesterol index values more than double along the OMZ transect and the below OMZ-seep and suggest faster degradation of cholesterol predominantly occurs with increasing oxygen concentrations at the SWI (Fig. 5b). More efficient degradation of cholesterol under oxic conditions has been suggested in experiments previously (Sun and Wakeham, 1998). However, our data contrast with those from the Peru margin OMZ, where anoxic sediments generally produced the highest ratios (McCaffrey et al., 1991). The reduction of cholesterol to cholestanol in anoxic surface sediments is generally a faster process than sterol degradation, and is the main 15 reason for high stanol/stenol ratios in anoxic waters and sediments (Nishimura and Koyama, 1977; Wakeham, 1989). However, the presence of an increasing trend along lateral oxygen gradients here suggests that the dominant process controlling this ratio is the degradation of cholesterol, so the ratio values reflect the faster degradation of cholesterol as opposed to cholestanol rather than the conversion of cholesterol to 20 cholestanol. The higher rate of stenol degradation versus stenol hydrogenation has

been suggested as one possibility for high stanol/stenol surface sediment values in an estuarine environment (Arzayus and Canuel, 2004). Therefore, in sediments with a steep lateral oxygen gradient, this ratio may be useful in determining the extent of sterol oxidation.



5.3.3 Dinosterol-based index

The dinosterol-based index increases by an order of magnitude along the OMZ transect and the below OMZ-seep (Fig. 5b), indicating faster degradation of dinosterol with increasing O₂ content at the SWI. It is unlikely that the index changes are a result of fluctuations in input from dinoflagellates, or other dinosterol-producing organisms (Volkman et al., 1993), as the trends are similar for the OMZ transect and the below OMZ-seep. Mouradian et al. (2007) indicate that inclusion of diagenetic products (Σ-dinflagellate lipids) of dinosterol provides evidence of preferential removal of dinosterol in oxic environments, as degradation of dinosterol would result in enrichments of its degradative products. Though dinostanone and dinosterone can be biosynthesized directly by dinoflagellates (e.g. Harvey et al., 1988; Leblond and Chapman, 2002; Chu et al., 2009), differences in input are again unlikely. Thus, we conclude that a dinosterol-based index using oxidative products of dinosterol degradation can be used to trace changes in oxygen content at the SWI.

15 5.3.4 G/P ratio

Dinoflagellate cysts are, in general, relatively resistant to degradation due to the structure of the cyst wall (e.g. Kokinos et al., 1998; Gélinas et al., 2001; de Leeuw et al., 2006; Zonneveld et al., 2008; Versteegh et al., 2011). However, it is known from laboratory (e.g. Dale, 1976; Hopkins and McCarthy, 2002) and field experiments (e.g. Kodrans-Nsiah et al., 2008) as well as natural sediments (e.g. Zonneveld et al., 1997, 2001, 2007; McCarthy et al., 2000) that P-cysts are more quickly oxidized relative to the G-cysts. In both the OMZ transect and below OMZ-seep, the G/P ratio increases by 75% and 66%, respectively. Thus, our data confirm the selective aerobic degradation of P-cysts and furthermore demonstrates a marked overprint along a meter-scale lateral evaluation and accient (below OMZ ecep). Province activities have acutioned against the

eral oxygen gradient (below OMZ-seep). Previous studies have cautioned against the use of this ratio because it oversimplifies the relationship between the heterotrophic taxa that indicate productivity and P-cysts (e.g. Dale and Fjellså, 1994). Our results



add another layer of caution as overprinting of the G/P ratio is likely to happen quickly with changes in oxygen content at the SWI. This overprint can be accounted for in some cases (Versteegh and Zonneveld, 2002) and, in areas of excellent preservation or when redox conditions remain stable over time, P-cysts are still valuable productivity proxies (e.g. Reichart and Brinkhuis, 2003). Therefore, the G/P ratio, based on the quantity of cysts, is also a potential proxy for tracing oxygen concentration changes at the SWI.

It is interesting to note that the dinosterol-based index and the G/P ratio demonstrated similar trends along the OMZ transect and the below OMZ-seep. There is some discrepancy between dinoflagellate cyst abundance and concentrations and dinosterol concentrations with studies either showing a weak (e.g. Marret and Scourse, 2002) or nonexistent (Pinturier-Geiss et al., 2002; Sangiorgi et al., 2005) correlation, which may be related to differences in the species that manufacture dinosterol as a membrane lipid and those that produce resting cysts (Boere et al., 2009). Our results agree with Mouradian et al. (2007) that inclusion of diagenetic transformation products of dinos-

¹⁵ Mouradian et al. (2007) that inclusion of diagenetic transformation products of dinosterol increases the likelihood that the dinosterol-based index and dinoflagellate cyst trends will be similar. This may then reflect a correlation that is based on diagenetic factors, rather than productivity.

5.4 Temperature proxy

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The overall small changes in the three GDGT-based indices and lack of any consistent trends (Fig. 5c) that could be attributed to the oxygen content at the SWI indicate that the GDGT distribution is not affected by selective aerobic degradation. Instead, it appears the GDGT signal is complicated by different factors in each transect as the TEX₈₆ values and resulting SSTs are variable. SSTs calculated from these indices do not consistently reflect the annual mean SST (26.7 °C) as recorded by Lornacini et al. (2010). TEX₈₆ values are higher than previously reported for the Arabian Sea (e.g. Schouten et al., 2002, 2004; Kim et al., 2008), and result in unrealistically high SSTs. The same is true for the TEX^H₈₆, which has shown to be suitable in subtropical



waters (Kim et al., 2010). The most similar calculated SST to the reported SST of the area is derived from the TEX^L₈₆, which has been shown to be appropriate in (sub) polar waters (Kim et al., 2010). The difference between the three indices is the absence of the crenarcheaol regioisomer (Cren iso.) in TEX^L₈₆, which suggests a deviation from its observed correlation with SST at low temperatures (Kim et al., 2010) and indicates that this index may not necessarily be limited to lower temperatures (Ho et al., 2011). The crenarchaeol regioisomer has been suggested to have a different source from the other GDGTs (see the discussion in Shah et al., 2008). It was pointed out in Shah et al. (2008) that this could have implications for the TEX₈₆ paleothermometer, which our data illustrate. However, the GDGT indices seem to show no clear trends that might suggest differential input of the crenarchaeol regioisomer or diagenetic alteration.

Complicating factors could include a strong seasonal signal (Wuchter et al., 2006; Leider et al., 2010), or sedimentary production in anoxic sediments (e.g. Pancost et al., 2001; Biddle et al., 2006; Lipp et al., 2008) so our data may reflect differential

- GDGT contributions from the water column (surface and subsurface; e.g. Huguet et al., 2007; Mollenhauer et al., 2008) as well as the seep environment. However, the lack of any consistency between OMZ-seep and below OMZ-seep would suggest that the two seep transect communities are heterogeneous. The recently proposed methane index (Zhang et al., 2011), suggests that in environments heavily impacted by methane
- ²⁰ hydrates, the contribution of GDGTs produced by methanotrophic archaea render the TEX₈₆ index unreliable. However, in our data, it appears that the crenarcheaol regioi-somer is the main source of variability as the TEX₈₆ produces the most realistic SSTs. Overall, we cannot determine which possible factors have the most influence but it seems that the three transects used in this study reflect heterogeneous input from the
- water column and the sediments. This suggests that, as discussed by Kim et al. (2010), there is a possibility of a significant effect from local conditions.



6 Conclusions

The OMZ and active methane seeps of the northeastern Arabian Sea present a natural laboratory to investigate proxy alteration as a result of changing oxygen content at the SWI. Our study investigated the effects of early selective aerobic degradation on
 ⁵ alteration, export production, and temperature proxies in surface samples collected along two lateral oxygen gradients. In order to constrain uncertainty regarding other environmental conditions, we utilized both the OMZ as well as methane seeps in oxic and anoxic bottom waters. The oxygen concentration at the SWI seemed to be the dominant controlling factor for most of the investigated proxy ratios, with the exception
 of the GDGT- and phytol-based indices, where localized environmental heterogeneity seems to be more important.

The HPA, API, and DOXI, previously used in sediment core studies as alteration proxies, appear to be quite sensitive to changing oxygen conditions at the SWI. These three proxies demonstrate sensitivity to changes in oxygen content at the SWI so that

- they may be used in combination in studies where OM composition or environmental heterogeneity is not as well constrained. The degradation of cholesterol at a welloxygenated SWI seems to be faster than the reductive transformation of cholesterol to cholestanol so that a stanol/stenol index may be a useful proxy for changes in oxygen content. Likewise, a degradation index based on dinosterol and its transformational
- ²⁰ products might be useful to show redox changes in surface sediments. Finally, the dinoflagellate cyst based G/P index may be a useful proxy for SWI oxygenation. Our data demonstrate that even a small, localized lateral oxygen gradient at the SWI is sufficient to induce selective OM degradation. This potential to quickly and significantly affect proxy signals provides further evidence that care must be taken in order to accurately interpret the sedimentary record.
- ²⁵ interpret the sedimentary record.



Supplementary material related to this article is available online at: http://www.biogeosciences-discuss.net/8/11359/2011/ bgd-8-11359-2011-supplement.pdf.

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 Table 1. Sample location descriptions and oxygen classification scheme.

Sample	Referred to	Latitude	Longitude	Water depth	Transect	Sample	*[O22]	Mn ²⁺	Oxygen	Based
	in text as	(N)	(E)	(m)		location	ml I ⁻¹	at SWI	classification	on
GeoB 12312	OMZ-anoxic	24 °53'	63°01′	655	OMZ	OMZ core	0.02	-	anoxic	CTD of the station,
GeoB 12321	OMZ-suboxic	24°46'	62°59'	1425	OMZ	OMZ lower transition	0.55	-	suboxic	O2 penetration depth,
GeoB 12331	OMZ-oxic	24°11′	62°46′	2830	OMZ	well below OMZ	2.55	-	oxic	Fe ²⁺
GeoB 12326-13	below OMZ-seep 1	24°11′	62°44′	2875	below OMZ seep	adjacent to gas orifice	-	yes	anoxic	Mn ²⁺ profile
GeoB 12326-9	below OMZ-seep 2	24°11′	62°44′	2875	below OMZ seep	50 cm from gas orifice, clam field	-	no	suboxic	Mn ²⁺ profile
GeoB 12326-7	below OMZ-seep 3	24°11′	62°44′	2875	below OMZ seep	1 m from gas orifice, ambient sediment border	2.55	-	oxic	CTD of the station
GeoB 12328-6	OMZ-seep 1	24 °50'	63°01′	1025	OMZ seep	bacterial mat	0.07	-	anoxic	
GeoB 12328-2	OMZ-seep 2	24°50'	63°01′	1025	OMZ seep	15 cm from mat, clam border	0.07	-	anoxic	CTD of the station
GeoB 12328-4	OMZ-seep 3	24°50′	63°01′	1025	OMZ seep	75 cm from central habitiat, clam field	0.07	-	anoxic	

* As measured by CTD profiling of bottom waters.

Table 2. Gonyaulacoid (G) and peridinioid and cysts of *Polykrikos* (P) dinoflagellate cyst species identified in this study.

P-cysts	G-cysts
Brigantedinium spp.	Bitectatodinium spongium
Cyst of <i>Diplopelta parva</i>	Impagidinium aculeatum
Cyst of Polykrikos kofoidii	Impagidinium paradoxum
Cyst of Protoperidinium americanum	Impagidinium patulum
Cyst of Protoperidinium monospinum	Impagidinium sphaericum
Cyst of Protoperidinium stellatum	<i>Impagidinium</i> spp.
<i>Dubridinium</i> spp.	Lingulodinium machaerophorum
Echinidinium aculeatum	Nematosphaeropsis labyrinthus
Echinidinium bispiniformum	Operculodinium centrocarpum
Echinidinium granulatum	Operculodinium israelianum
Echinidinium transparantum	Operculodinium longispinigerum
Echinidinium delicatum	Pentapharsodinium dalei
<i>Echinidinium</i> spp.	Polysphaeridium zoharyi
Leipokatium invisitatum	Spiniferites membranaceous
Lejeunacysta oliva	Spiniferites mirabilis
Lejeunacysta sabrina	Spiniferites pachydermus
<i>Lejeunacysta</i> spp.	Spiniferites ramosus
Quinquecuspis concreta	<i>Spiniferites</i> spp.
Selenopemphix nephroides	
Selenopemphix quanta	
Stelladinium robustum	
Trinovantedinium applanatum	
Votadinium calvum	
Xandarodinium xanthum	



Table 3. Proxy definition.

Proxy	Definition	Components	Source	Source references	Proxy represents	Proxy references	
Alteration indices							
HPA	$[\sum C_{24-28} \text{ even-OH}]/ [\sum C_{24-28} \text{ even-OH}]$	long chain even n-alcohols,	Higher plant waxes	Eglinton and Hamilton (1967), Fukushima and Ishiwatari (1984)	Preservation	Westerhausen et al. (1993), Yamamoto et al. (2008)	
	+ [∑C ₂₇₋₃₁ odd <i>n</i> -alkanes]	odd n-alkanes					
API	[C ₂₆ -OH]/[C ₂₆ -OH] + [C ₂₉ <i>n</i> -alkane]	C ₂₆ <i>n</i> -alcohol, C ₂₉ <i>n</i> -alkane	Same as HPA		Bottom water oxygenation	Cacho et al. (2000), Martrat et al. (2007), Versteegh et al. (2010)	
DOXI	[1,15-keto-ol]/ [1,15-diol] + [1,15-keto-ol]	C ₃₀ and C ₃₂ 1,15 alkyl diols, keto-ols	Eustigmatophytes, unknown for keto-ols	de Leeuw et al. (1981), Versteegh et al. (1997)	Preservation	Ferreira et al. (2001), Versteegh et al. (2010)	
Export production indices							
Phytol-based							
index'	[pristane] + [phytane]/ [phytol]	Phytol, phytane, pristane	Chlorophyll-a	Volkman and Maxwell (1986), Didyk et al. (1978)	Preservation	modified from Didyk et al. (1978), This study	
index ²	[phytane]/[phytol]						
index ³	[pristane /[phytol]	Chalastaral shalastanal	Fukanistan	Parrat at al. (1005)	Photic zono productivity	Nishimura and Kayama (1077)	
Stanorstenor	[cholestanol/[cholesterol]	Cholesterol, cholestanol	Eukaryotes	Volkman et al. (1999), Hudson et al. (2001)	stenol reduction	McCaffrey etal. (1991)	
Dinosterol-based	[dinosterone] + [dinostanone] + [dinosterane]/[dinosterol]	dinosterol, dinosterone, dinostanone, dinosterane	Dinoflagellates	Boon et al. (1979), Robinson et al. (1984), Volkman et al. (1999)	Photic zone productivity, oxic degradation	Mouradian et al. (2007), This study	
G/P	[gonyaulacoid cysts]/[peridinioid cysts] + [cysts of <i>Polykrikos</i>]	Organic-walled, non-motile resting cysts	Dinoflagellates	Fensome et al. (1993)	Photic zone parameters, redox changes	McCarthy et al. (2000), Mudie and Rochon (2001)	
Temperature indices*							
TEX ₈₆	[GDGT-2]+[GDGT-3] + [Cren. Iso]/ [GDGT-1] + [GDGT-2] + [GDGT-3] + [Cren. Iso]	Glycerol dibiphanyl glycerol tetraethers	marine Crenarchaeota	Karner et al. (2001) Schouten et al. (2002)	Sea surface temperature	Schouten et al. (2002)	
TEX ^L ₈₆	log [GDGT-2] / [GDGT-1] + [GDGT-2] + [GDGT-3]					Kim et al. (2010)	
TEX ^H ₈₆	log[GDGT-2] + [GDGT-3] + [Cren. Iso] [GDGT-1] + [GDGT-2] + [GDGT-3] + [Cren. Iso]/						

* Numbers in the temperature index definitions refer to the number of cyclopentane moieties in the GDGT structure. Cren. Iso. refers to the crenarchaeol regioisomer.

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Table 4. Proxy values of the calculated indices along the three studied transects.

		OMZ transect		Below OMZ seep				OMZ seep			
	OMZ-anoxic	OMZ-suboxic	OMZ-oxic	below OMZ-seep 1	below OMZ-seep 2	below OMZ-seep 3	OMZ-seep 1	OMZ-seep 2	OMZ-seep 3		
	(anoxic)	(SUDOXIC)	(OXIC)	(anoxic)	(suboxic)	(UXIC)	(anoxic)	(anoxic)	(anoxic)		
Alteration indices											
HPA	0.58	0.52	0.42	0.67	0.65	0.55	0.68	0.73	0.6		
API	0.47	0.39	0.18	0.56	0.49	0.37	0.61	0.72	0.67		
DOXI											
C ₃₀ isomer	0.12	0.24	0.53	0.26	0.37	0.41	0.28	0.26	0.25		
C ₃₂ isomer	0.14	0.22	n.m.	0.27	n.m.	n.m.	0.28	0.26	n.m.		
Export production indices											
Phytol-based											
index ¹	0.28	0.24	0.25	0.51	0.25	0.25	1.26	0.62	0.47		
index ²	0.16	0.08	0.15	0.2	0.1	0.11	0.28	0.19	0.15		
index ³	0.11	0.16	0.15	0.32	0.2	0.15	0.99	0.43	0.32		
Stanol/stenol	0.23	0.24	0.82	0.37	0.59	0.75	0.54	0.61	0.4		
Dinosterol-based	0.02	0.05	0.24	0.03	0.21	0.26	0.18	0.09	0.06		
G/P	0.13	0.31	0.52	0.15	0.3	0.44	0.17	0.18	0.18		
Temperature indices*											
TEX ₈₆ ^a	0.75 (34.6°C)	0.75 (34.7 °C)	0.72 (31.7 °C)	0.76 (35.6 °C)	0.73 (32.7°C)	0.69 (29.2°C)	0.72 (31.7°C)	0.71 (31.0°C)	0.75 (34.9 °C)		
TEX ₈₆	-0.29 (27.5°C)	-0.32 (25.5 °C)	-0.29 (27.2°C)	-0.3 (26.8 °C)	-0.33 (24.3°C)	-0.33 (24.6°C)	-0.3 (26.7°C)	-0.29 (27.0°C)	-0.28 (28.3 °C)		
TEX ^{H c}	-0.12 (30.1 °C)	-0.12 (30.2 °C)	-0.15 (28.6 °C)	-0.12 (30.6 °C)	-0.14 (29.2°C)	-0.16 (27.4°C)	-0.15 (28.7 °C)	–0.15 (28.3 °C)	-0.12 (30.3 °C)		

Definitions of each of the listed indices can be found in Table 3.

* Index values are followed in brackets by the calculated SST with that value. All calculations are after Kim et al. (2010).

^a SST = $81.5 \times TEX_{86} - 26.6$, ^b SST = $67.5 \times TEX_{86}^{L} + 46.9$,

^c SST = $68.4 \times TEX_{86}^{H} + 38.6$.





Fig. 1. Map showing (a) sample locations (b) water column oxygen concentrations, as measured by CTD profiling, in relation to sample locations. Circles indicate multi-cores and stars denote push core locations.

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Interactive Discussion

a) OMZ-seep transect (GeoB 12328)

OMZ-seep 1

OMZ-seep 2





b) below OMZ-seep transect (GeoB 12326)

below OMZ-seep 1

below OMZ-seep 2





Fig. 2. High resolution photographs taken by the ROV MARUM-QUEST 4000 of the individual push cores retrieved along the two seep transects **(a)** OMZ-seep (GeoB 12328), and specific samples OMZ-seep 1 (GeoB 12328-6), OMZ-seep 2 (GeoB 12328-2), and OMZ-seep 3 (GeoB 12328-4), **(b)** GeoB 12326 (below OMZ-seep), and specific samples below OMZ-seep 1 (GeoB 12326-13), below OMZ-seep 2 (GeoB 12326-9), and below OMZ-seep 3 (GeoB 12326-7).







Fig. 4. Mn^{2+} concentrations (μM) from two replicate push cores of below OMZ-seep 1 and 2.





Fig. 5. Proxy ratio trends along the three studied transects of (a) alteration indices, (b) export production proxies, and (c) temperature proxies.

